

EVALUATION OF ORGANIC WASTES AS SUBSTRATES FOR REARING *ZOPHOBAS MORIO*, *TENEBRIO MOLITOR*, AND *ACHETA DOMESTICUS* LARVAE AS ALTERNATIVE FEED SUPPLEMENTS

RICHÁRD PINTÉR – GYÖRGY FEKETE – ZSOLT ISTVÁN VARGA –
CSABA GYURICZA – LÁSZLÓ ALEKSZA

Abstract

*Studies have focused on identifying combinations of insects and organic waste to optimise bio-conversion. Here, the effects of different diets (10% chicken feed complemented with 90% vegetable waste, garden waste, cattle manure, or horse manure) on growth and survival rates and nutritional value of *Zophobas morio* and *Tenebrio molitor* larvae, and *Acheta domesticus* were investigated. Compared with chicken feed, organic waste decreased the individual larval weight, although green waste showed fewer negative effects than the manure. The macronutrient concentrations in garden waste were moderate compared with chicken feed, and vegetable waste was the poorest diet in terms of nutrient concentration. There was no difference in weight between larvae reared on garden waste and those reared on vegetable waste. *Tenebrio molitor* and *A. domesticus* showed the maximum growth rates at 71–101 and 36–66 days of age at 22.5 ± 2.5 °C, respectively. *Acheta domesticus* was rich in proteins, whereas *Z. morio* and *T. molitor* were rich in fat. Feeding nutrient-poor diets resulted in a lower protein and a higher fat concentration in the larvae.*

Keywords: larva, growth, survival rates, nutritional value, feed

***ZOPHOBAS MORIO*, *TENEBRIO MOLITOR* ÉS *ACHETA DOMESTICUS* LÁRVÁK TENYÉSZTÉSÉNEK ÉRTÉKELÉSE SZERVES HULLADÉKOKBÓL ÁLLÓ SZUBSZTRÁTOK, MINT ALTERNATÍV TAKARMÁNYKIEGÉSZÍTŐK HOZZÁADÁSÁVAL**

Összefoglalás

*A tanulmány célja különböző takarmányok hatásának vizsgálata volt *Zophobas morio* és *Tenebrio molitor* lárvák, valamint az *Acheta domesticus* növekedési és túlélési arányára, valamint beltartalmi jellemzőjükre. A kontroll csoportok takarmányként csak csirketápot kaptak, a vizsgált takarmány keverékek 10% csirketáp és 90% szerves hulladék keverékéből álltak (növényi hulladék, zöldség hulladék, szarvasmarha trágya és lótrágya). A csirketakarmánnyal összehasonlítva a szerves hulladék csökkentette az egyes lárvák tömegét, bár a zöldhulladék kevesebb negatív hatást mutatott, mint a trágya. A kerti hulladék makrotápanyag-koncentrációja mérsékelt volt, összehasonlítva a csirketakarmánnyal, és a növényi hulladék volt a tápanyagkoncentráció szempontjából a legszegényebb étrend. Tömegben nem volt különbség a kerti hulladékon és a növényi hulladékon nevelt rovarok között. A *Tenebrio molitor* és az *A. domesticus* a maximális növekedési sebességet 71-101, illetve 36-66 napos korban $22,5 \pm 2,5$ °C-on mutatta. Eredményeink megerősítették, hogy a vegyes*

növényi hulladék, több faj zöld biomassájú kerti hulladék, illetve a szarvasmarha- és lótrágya magas aránya nem tekinthető optimális tenyésztési szubsztrátumnak az *A. domesticus*, *T. molitor* és *Z. morio* lárvák termesztésére. A tápanyag-szegény étrend etetése alacsonyabb fehérjetartalmat és magasabb zsírkoncentrációt eredményezett a lárvákban.

Kulcsszavak: *lárva, növekedés, túlélési arány, tápérték, takarmány*

JEL code: Q53

Introduction

Edible insects are of high nutritional value, and they can be used to partially replace protein ingredients in compound feeds for common livestock and aquaculture. Furthermore, insects of different compositions can be considered an ingredient in feed formulations (BENZERTIHA et al., 2020). The most common commercial insect species are the yellow mealworm (*Tenebrio molitor* L.), super worm (*Zophobas morio* Fab.), housefly (*Musca domestica* L.), black soldier fly (*Hermetia illucens* L.), house cricket (*Acheta domesticus* L.), and greater wax moth (*Galleria mellonella* L.) (ORTIZ et al., 2016; VARELAS, 2019). The composition of insect larvae varies with species, and it may be either species-specific or modified by their diet.

For large-scale production of insect-based feed supplements, the cost of rearing substrates is a key issue. The major expenses in insect rearing are related to procuring raw materials used for feeding the insects (CADINU et al., 2020) and maintaining elevated temperature for most species (MORALES-RAMOS et al., 2018). In the European Union, edible insects can be fed only materials of vegetable origin and certain materials of animal origin. However, they can grow efficiently on bio-waste and by-products such as restaurant and household waste, slaughterhouse by-products, animal manure, and gardening waste (LÄHTEENMÄKI-UUTELA – GRMELOVÁ, 2016). Thus, feed regulations should be revised to encompass these alternatives in the circular economy; these raw materials should be authorised for utilisation. Studies have focused on rearing insects on low-value substrates, such as organic wastes and by-products, and indicated their advantages and limitations. VARELAS and LANGTON (2017) reviewed the potential of forest biomass by-products for rearing edible insects and provided examples of unbalanced feeding and its negative consequences on growth and nutritional values. KIM et al. (2014) investigated the effect of agricultural waste as a feed for *T. molitor* by replacing mushroom substrates with wheat bran feed. Although the larvae thus reared were lighter and required longer development periods than the control group larvae, their survival rate was similar to that of larvae reared on mushroom substrates.

Oonicx et al. (OONICX et al., 2015) fed by-products of the food industry to *T. molitor* and *A. domesticus*, identified diets that can be as efficiently converted by the larvae as pigs, and concluded that when fed optimal diets, larvae can convert their feed as efficiently as poultry, after correcting to the edible portion. Especially for nitrogen efficiency, their performance was higher than that of conventional production animals. Diet affected the survival and development period of *T. molitor*; furthermore, feeding carrots increased the dry matter content and nitrogen efficiency, and decreased the development period. LUNDY and PARRELLA (2015) measured the biomass output and feed conversion ratio of *A. domesticus* reared on diets ranging from grain-based to highly cellulosic diets, and concluded that the nitrogen concentration, N-to-acid detergent fibre concentration ratio, and crude fat concentration explained most of the variabilities among feed treatments. However, the mortality rate of crickets fed minimally processed, municipal-scale food waste and diets composed largely of straw was >99% without reaching a harvestable size. Some studies have revealed that low-value diets may be effective even at temperatures lower than the optimal range (ADÁMKOVÁ et al., 2017a; BOOTH – KIDDELL, 2007).

On the basis of the above findings, the following can be inferred: the growth of larvae can be maximised by nutrition; chemical composition can be functionally pre-designed; and low-value, organic by-products and waste have great potential as substrates for increasing economic feasibility in insect rearing. Larval performance and waste biotransformation depend on the chemical composition of the organic by-products selected. Therefore, the objective of the current study was to compare the effects of two types of green waste and two types of manure as rearing substrate on the larval performance of three edible species, *T. molitor*, *Z. morio*, and *A. domesticus*, under the same environmental conditions, during their intensive growing period. The substrates were evaluated for their effect on the growth, nutritional values, and mortality of the larvae.

Materials and Methods

Insects and Diets

For the study, the superworm (*Zophobas morio*, *Coleoptera: Tenebrionidae*), yellow mealworm (*Tenebrio molitor*, *Coleoptera: Tenebrionidae*), and house cricket (*Acheta domesticus*, *Orthoptera: Gryllidae*) were selected among the seven commercially available insect species for feeding animals in the EU. *Acheta domesticus* and *T. molitor* are among the most intensively investigated species, whereas *Z. morio* has not been extensively studied, although it is also mass-produced. The insects were procured from BUGS-WORLD Ltd. (Tiszaékéske, Hungary).

The organic wastes were:

- Vegetable waste (mixed peels of 10% onion, 25% potato, 25% sweet potato, 30% carrot, and 10% cucumber, with a total water content of 91.4%);
- Green garden waste with grass (50% Poaceae species and other common weeds, 25% tree leaves, and 25% branches (Populus, Salix, Pinus, and Corylus species), and a mixture of stone fruits, and other ornamental plant parts, with a water content of 36.2%);
- 55% cattle manure with faeces and urine, and 45% cereal straw with a water content of 45.7%;
- 35% horse manure with faeces and urine, and 65% cereal straw with a water content of 28.3%.

For the experiments, the larvae were fed diets (henceforth called substrates) containing 90% of the given organic waste and 10% of chicken feed. The green waste was chopped into 2–4-cm long pieces and manures were broken into small pieces. Mashed chicken feed, produced by VITAFORT Plc. (Dabas, Hungary) for intensive broiler breeding (13.05% water, 0.80% lysine, and 0.30% methionine) was mixed with the substrates and was used as a control. Chicken feed was chosen considering both industrial practice and recommendations and previous study results (MIECH et al., 2016; RUMBOS et al., 2020). Preliminary tests showed a high mortality of insects with all combinations of the selected organic waste in pure form. However, providing a limited amount of balanced feed improved larval survival; 10% chicken feed supplementation generally resulted in a low mortality rate.

Experimental Design

Before starting the feeding experiments, the newly hatched larvae of *T. molitor* and *Z. morio* and those of *A. domesticus* were reared on chicken feed, fresh carrots, and cucumber (70%, 20%, and 10%, respectively) for 56 and 21 days, respectively, when they started to grow intensively. The diet provided during this period ensured that the larvae were in good condition at the start of the experiment. Air humidity was maintained at 60% ± 4% for *T. molitor* and *Z.*

morio and 80%, 70%, and 60% for *A. domesticus* during the subsequent weeks to provide optimal conditions for healthy insect development. In the rearing environment, the temperature was 22.5 ± 2.5 °C and humidity was $60\% \pm 4\%$, with a 12:12-h light/dark cycle. We focused on the intensive growing period of the selected species under the given rearing conditions. The experiments lasted for 45 days with all three insects. The initial number of larvae was 100 in each trial, with three replications. The larvae were fed ad libitum throughout the experimental period. Fresh substrates of weight equivalent to $25\times$ the net weight of live larvae were added, and the residues and excreta were removed on days 15 and 30 when recording the weight of the larvae. The size of the plastic box (width \times length \times height) for *T. molitor* and *Z. morio* was $30 \times 38 \times 10$ cm, and that for *A. domesticus* was $18.1 \times 25.6 \times 13.6$ cm; egg cartons with a surface area of approximately 1800 cm² were used in each trial.

Measured Parameters

Mortality, growth, and weight of live larvae were recorded on days 15, 30, and 45. After maintaining the larvae at 4 °C for 60 min, they were separated from the remaining substrate and excreta with a spatula in the first period (later by sieving with the mesh of size 2 mm), and then further separated as dead and alive. Only live individuals from each experimental unit were considered when recording. The larvae were weighed using a pre-calibrated KERN ABT 320-4NM analytical balance of 0.1 g weighing accuracy, with a measuring range of 10 mg–320 g. The nutritional composition of the larvae, including crude protein, crude fat, fibre, ash, and energy, was measured after 45 days of the experiment. The substrates were analysed for the total organic content, and total nitrogen, protein, carbohydrate, fat, total phosphorous, potassium, and calcium concentrations. All chemicals and reagents were of analytical grade. Total nitrogen concentration was determined using the Kjeldahl method according to the standard ISO 5983-1:2005 method for animal feedstuff. Crude protein concentration (P) was calculated using Equation 1

$$P = \text{total Kjeldahl nitrogen} \times CF, \quad (1)$$

Where CF is the conversion factor, which is 4.76 for the larvae and 6.25 for the substrates. JANSSEN et al. (2017) proved that nonprotein N in insects leads to an overestimation of protein concentration. They reported comparable CF values among larvae belonging to different orders; the CF for *T. molitor* was 4.76 ± 0.09 . Crude fat concentration in the substrates and the larvae was determined using the standard ISO 11085:2015 method for cereals, cereal-based products, and animal feedstuffs using an automated extractor (VELP Scientific, Randall). Carbohydrate concentration was determined using a spectrophotometer (Hach DR6000), according to the method of DUBOIS et al. (1956). Fibre concentration was measured according to the method of BORDEREAU and ANDERSEN (1978) for termite species. Total ash concentration was determined using the standard ISO 936:2000 method for meat and meat products. Gross calorific value was determined using the standard ISO 9831:1998 method with a Parr 6400 automatic isoperibol oxygen bomb calorimeter. Substrate extracts were prepared using the microwave-assisted digestion method with 2 mol/dm³ nitric acid and 30% m/m hydrogen peroxide, on a Milestone MLS 1200 Mega high-performance microwave digestion unit. The concentration of total organic carbon (TOC), phosphorous, and potassium was determined using the Hach DR6000 spectrophotometer following the LCK 381, LCK 350, and LCK 228 tests, respectively. Calcium concentration in the extracts was measured using a Jenway PFP7 type low-temperature, single-channel, flame photometer.

Data Analyses

The changes in mortality and individual weight of the larvae over time were analysed using a repeated-measures ANOVA, where the larval characteristics were considered the dependent variables, whereas species and rearing substrates were considered the independent variables. The dependent variables were continuous, whereas the responding ones were categorical. The four-time points (days 0, 15, 30, and 45) were chosen as the repeated measure factor. Time, species, and substrate interactions were considered statistically different at the significance level of 5%. When the F-test results were significant, Duncan's new multiple range tests were used for post-hoc comparison of the differences between the pairs of means at a level of $\alpha = 0.05$. Statistical computing was carried out using R software. Data were analysed using R version 3.6.3 (2020-02-29).

Results

Effect of Substrates on Larval Weight

An increase in the weight of individual larvae was monitored for each species and substrate. There were significant differences between the weights recorded every 15 days for all diets and species (Figure 1). The Duncan test differentiated the three homogenous groups ($p < 0.10$). There were no significant differences in weight among larvae of the three species and between the experimental diets.

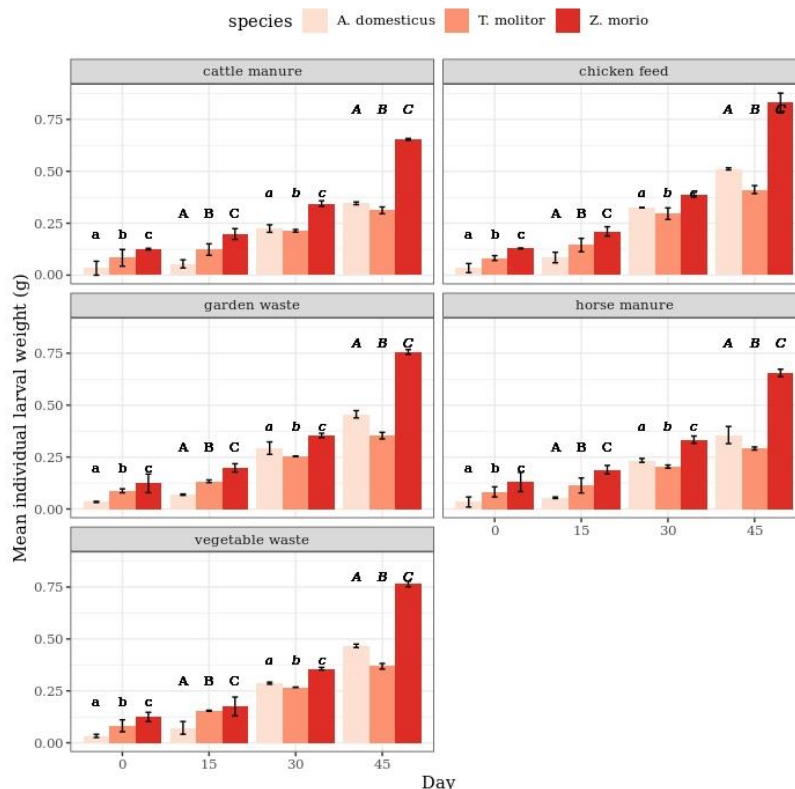


Figure 1. Mean individual larval weight of *T. molitor* and *Z. morio* at the age of 56–101 days, and *A. domesticus* at the age of 21–66 days, by substrates (the bars indicate the 95% confidence interval). Different letters indicate statistically significant differences between only the species (lowercase and uppercase does not indicate differences, just to visually separate days)

Considering the increase in larval weight during the 45-day experiment period, regardless of the species and rearing substrate, *Z. morio* showed the highest growth rate, followed by *A. domesticus*, and *T. molitor*. The difference in the growth rate between the latter two species changed the order of individual weight between days 15 and 30; it is noteworthy that the initial ages of the larvae were different (21 and 56 days), respectively (Figure 2).

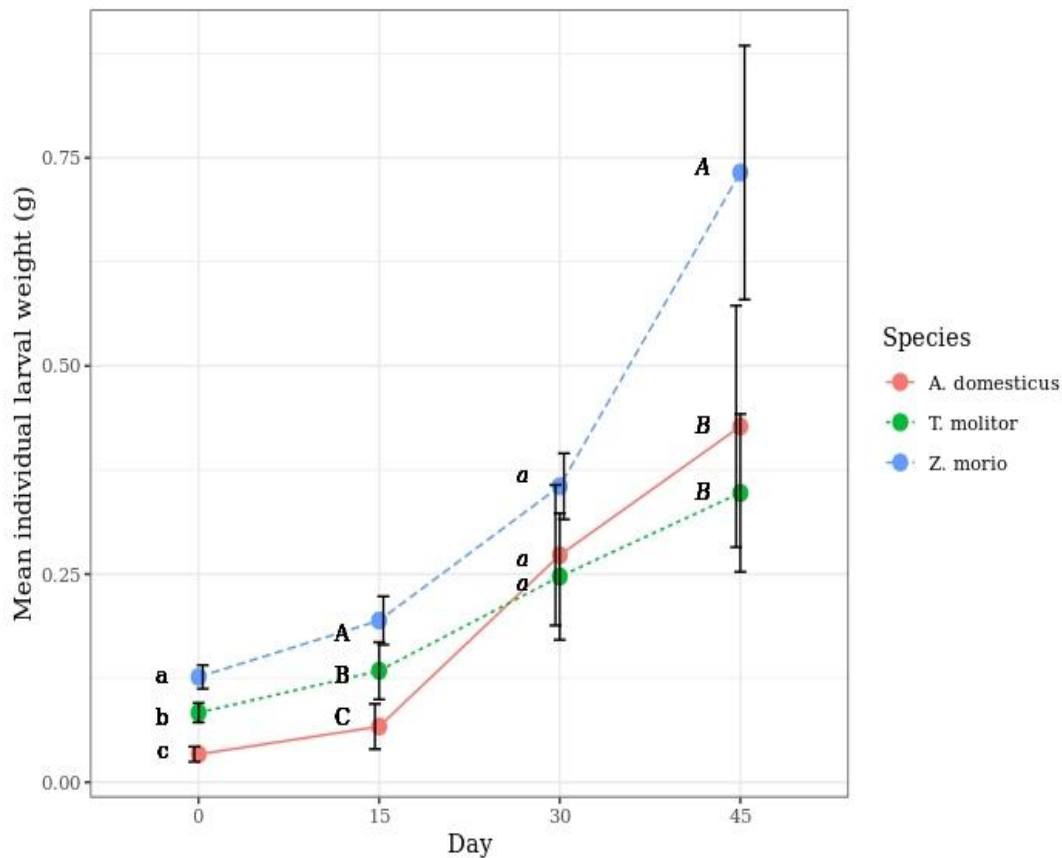


Figure 2. Mean individual larval weight (common mean of larval weight reared on different substrates) of *T. molitor*, *Z. morio*, and *A. domesticus* when reared on the different substrates, by time (the bars indicate the 95% confidence interval). Different letters show statistically significant differences among the species.

A comparison between the two species belonging to the family Tenebrionidae revealed that *Z. morio* was significantly heavier than *T. molitor* at the age of 56 days (1st day of experiment) ($\alpha = 0.05$), and this difference increased with time. *Acheta domesticus* larvae were the lightest, but they showed the highest relative increase in individual weight. However, during the experimental period of 45 days, the rate of growth with time, expressed as a rate of change $[(mt/mt-1) \times 100]$, increased for *Z. morio*, and started to decrease between days 30 and 45 for *T. molitor*, and peaked between days 15 and 45 for *A. domesticus* (Figure 3).

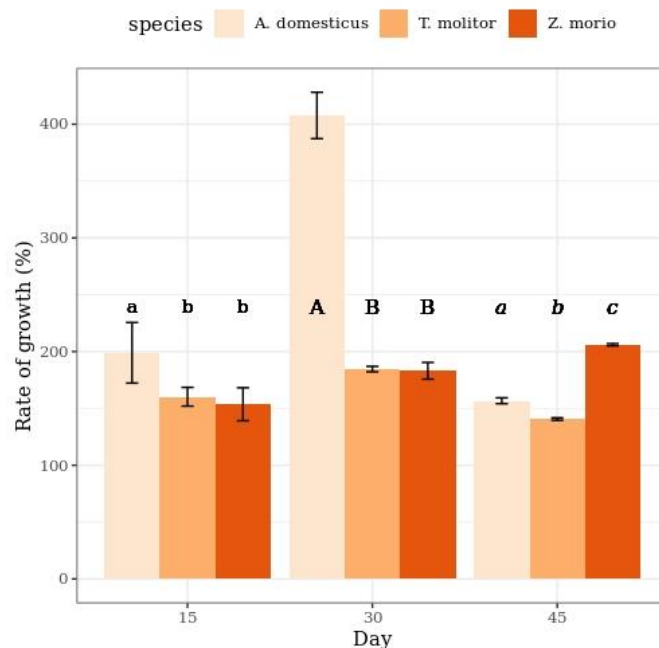


Figure 3. Rate growth of *T. molitor* and *Z. morio* larvae and *A. domesticus* when reared on the different substrates (the bars indicate the 95% confidence interval). Different letters show statistically significant differences among the species (calculated from common mean of larval weight reared on different substrates)

Nutritional Compositions of the Substrates

The nutrient composition of the substrates used in the experiment is presented in Table 1. The mean dietary ranges for different components were as follows: 9.43–30.08% TOC, 0.16–1.28% N, 27.88–212.80 g/kg protein, 13.35–125.70 g/kg carbohydrate, 2.34–5.58 g/kg fat, 0.10–0.43% K, and 0.79–1.38% Ca. Chicken feed had the highest total nitrogen, protein, and carbohydrate concentrations. The carbohydrate concentration of vegetable waste was comparable with that of other wastes. Fat concentration in the chicken feed was not significantly different from that in vegetable waste and cattle manure, and it was lower than that in garden waste and horse manure. The phosphorous concentration was different in all substrates, whereas the potassium concentration did not differ between the cattle and horse manure. The chicken feed contained the highest concentration of protein, nitrogen, carbohydrate, and phosphorous compared with the other four substrates. The fat and carbon concentrations were relatively higher in horse manure, but they were comparable between cattle manure and garden waste. Vegetable waste had the lowest concentration of fat and carbon.

Table 1. Nutrient composition of the substrates used as diets for rearing *Zophobas morio*, *Tenebrio molitor*, and *Acheta domesticus* larvae.

Components	CF	CF/VW 1:9	CF/GW 1:9	CF/CM 1:9	CF/HM 1:9
Total organic carbon (C%)	19.74 ± 3.38 ^b	9.43 ± 1.71 ^d	16.63 ± 2.25 ^c	19.66 ± 1.70 ^b	30.08 ± 2.16 ^a
Total nitrogen (N%)	1.28 ± 0.12 ^a	0.16 ± 0.01 ^d	0.60 ± 0.05 ^b	0.37 ± 0.03 ^c	0.30 ± 0.03 ^c
Protein (g/kg)	212.8 ± 8.31 ^a	27.88 ± 4.78 ^e	76.18 ± 7.74 ^b	39.65 ± 7.11 ^d	50.33 ± 5.84 ^e
Carbohydrate (g/kg)	125.7 ± 1.74 ^a	19.25 ± 0.70 ^b	14.01 ± 1.03 ^d	13.35 ± 0.71 ^d	16.90 ± 0.93 ^c
Fat (g/kg)	3.22 ± 1.08 ^c	2.34 ± 0.99 ^c	4.24 ± 0.70 ^b	3.19 ± 0.64 ^c	5.58 ± 0.60 ^a

Phosphorous (P%)	0.43 ± 0.02 ^a	0.10 ± 0.01 ^e	0.18 ± 0.02 ^c	0.16 ± 0.01 ^d	0.21 ± 0.02 ^b
Potassium (K%)	0.49 ± 0.04 ^b	0.10 ± 0.05 ^d	0.64 ± 0.06 ^a	0.37 ± 0.04 ^c	0.42 ± 0.05 ^c
Calcium (Ca%)	1.02 ± 0.02 ^c	0.97 ± 0.07 ^c	1.38 ± 0.03 ^a	1.15 ± 0.06 ^b	0.79 ± 0.03 ^d

Abbreviations: CF: chicken feed (control), VW: vegetable waste, GW: garden waste, CM: cattle manure, HM: horse manure; 1:9 is the ratio of CF and the given organic waste. The values are presented as mean ± SD in dry weight %, $n = 6$. Means within a row with the same letter are not significantly different.

Nutrient Value of the Larvae

The nutrient composition of the larval species, evaluated at the end of the experiments, showed differences with the diets (Table 2).

The crude protein concentration in *A. domesticus* was 57.8% when fed cattle manure, followed by 56.4% when fed horse manure; the protein concentration was the highest, that is, 67%, when fed chicken feed. The crude fat concentration was 14.4–19.4%, with the lowest and highest concentrations in larvae-fed chicken feed and horse manure, respectively. There were no significant differences between the larvae-fed manure and those fed garden waste. The fibre concentration ranged from 15.7–19.2%, with the lowest and highest concentrations in larvae fed chicken feed and manures, respectively. The ash concentration was the highest when the larvae were fed garden waste (6.2%), whereas it was the lowest when the larvae were fed the other diets. The larva- fed chicken feed presented the lowest energy level, whereas that fed manure exhibited the highest energy level, with a range of 17.4–18.8 MJ/kg. The crude protein concentration of *T. molitor* was lower, whereas its crude fat concentration was higher than those of *A. domesticus*, with a range of 37.9–47.2% and 43.1–47.5%, respectively. The fibre and ash concentrations were lower in *T. molitor*, whose fibre concentration was half of that of *A. domesticus*. Considering the differences in the nutrient composition of *T. molitor*, rearing the larvae on chicken feed resulted in the highest crude protein and the lowest crude fat concentrations, whereas rearing on horse manure resulted in the highest fat and the lowest protein concentration. Furthermore, the fibre and ash concentrations of the larvae were the lowest when reared on chicken feed, whereas they were the highest when the larvae were reared on the manures. *Zophobas morio* was comparable to *T. molitor* in terms of protein, fat, and ash concentrations. The order of change was similar, with the larvae reared on manure presenting the lowest protein and highest fat concentrations. Only the fibre concentration differed between the diets, wherein garden waste resulted in the highest fibre concentration, whereas horse manure resulted in a low fibre concentration compared with chicken feed. The mean ash concentration of *A. domesticus* was higher and less variable than that of *T. molitor* and *Z. morio*. The ash concentration of *A. domesticus* was independent of the substrate composition, but the manure diet increased and the green waste diet decreased the ash concentration in *T. molitor* and *Z. morio*.

Table 2: Analysed nutrient composition of *Zophobas morio*, *Tenebrio molitor*, and *Acheta domesticus* larvae by diet (the values are presented as mean ± SD in dry weight %, $n = 3$).

Components	CF	CF/VW 1:9	CF/GW 1:9	CF/CM 1:9	CF/HM 1:9
<i>A. domesticus</i>					
Crude protein (g/kg)	67.25 ± 0.10 ^a	61.20 ± 0.57 ^c	65.30 ± 0.50 ^b	57.80 ± 0.10 ^d	56.40 ± 0.40 ^e
Crude fat (g/kg)	14.41 ± 0.03 ^c	17.10 ± 0.65 ^b	19.30 ± 0.53 ^a	18.60 ± 0.29 ^a	19.40 ± 0.43 ^a
Fibre (g/kg)	15.72 ± 0.03 ^c	17.50 ± 0.66 ^b	17.83 ± 0.38 ^b	18.60 ± 0.29 ^a	19.20 ± 0.37 ^a
Ash (g/kg)	4.80 ± 0.04 ^b	5.40 ± 0.54 ^b	6.20 ± 0.39 ^a	5.20 ± 0.32 ^b	4.98 ± 0.32 ^b
Energy (MJ/kg)	17.35 ± 0.10 ^e	18.22 ± 0.01 ^d	18.27 ± 0.02 ^c	18.64 ± 0.01 ^b	18.78 ± 0.02 ^a

<i>T. molitor</i>					
Crude protein (g/kg)	47.18 ± 0.04 ^a	46.30 ± 0.47 ^b	42.30 ± 0.27 ^c	38.92 ± 0.48 ^d	37.90 ± 0.29 ^e
Crude fat (g/kg)	43.08 ± 0.05 ^e	43.30 ± 0.24 ^d	45.20 ± 0.43 ^c	46.70 ± 0.31 ^b	47.50 ± 0.36 ^a
Fibre (g/kg)	7.44 ± 0.02 ^d	8.01 ± 0.37 ^c	8.90 ± 0.41 ^b	9.50 ± 0.23 ^{ab}	9.30 ± 0.16 ^a
Ash (g/kg)	3.08 ± 0.05 ^b	3.01 ± 0.39 ^b	3.20 ± 0.38 ^b	4.85 ± 0.21 ^a	5.30 ± 0.33 ^a
Energy (MJ/kg)	24.17 ± 0.01 ^d	24.39 ± 0.02 ^c	24.59 ± 0.01 ^b	24.68 ± 0.01 ^a	24.60 ± 0.02 ^b
<i>Z. morio</i>					
Crude protein (g/kg)	46.79 ± 1.03 ^a	45.70 ± 0.48 ^b	41.20 ± 0.31 ^c	39.40 ± 0.34 ^d	38.70 ± 0.39 ^d
Crude fat (g/kg)	42.04 ± 0.74 ^d	43.20 ± 0.31 ^c	44.30 ± 0.37 ^b	45.70 ± 0.41 ^a	46.30 ± 0.42 ^a
Fibre (g/kg)	9.26 ± 0.04 ^c	9.43 ± 0.30 ^c	11.30 ± 0.11 ^a	10.20 ± 0.48 ^b	9.32 ± 0.20 ^c
Ash (g/kg)	2.61 ± 0.03 ^b	2.89 ± 0.31 ^b	3.01 ± 0.15 ^b	4.70 ± 0.34 ^a	4.89 ± 0.27 ^a
Energy (MJ/kg)	24.10 ± 0.02 ^d	24.43 ± 0.01 ^b	24.38 ± 0.01 ^c	24.75 ± 0.01 ^a	24.75 ± 0.01 ^a

Abbreviations: CF: chicken feed (control), VW: vegetable waste, GW: garden waste, CM: cattle manure, HM: horse manure; 1:9 is the ratio of CF and the given organic waste. Means within a row with the same letter are not significantly different.

Evaluation of the Effect of Substrates and Rearing Time on the Survival of Larvae

After 45 days of rearing, the highest recorded mortality of *Z. morio* and *T. molitor* larvae was 6.67% ± 0.58% and 2.76% ± 0.58%, respectively. At the end of the experiment, the mortality rate of *A. domesticus* fed chicken feed was 5.30% ± 1.53%. However, after 45 days of rearing, the percentage of live *A. domesticus* larvae was 77.67% ± 1.53% for vegetable waste, 68.00% ± 4.36% for garden waste, 64.33% ± 1.53% for horse manure, and 54.67% ± 1.53% for cattle manure. When fed green garden waste and vegetables, there was a considerable drop in the number of live larvae only on day 45. The results of Duncan's multiple range test showed significant differences ($\alpha = 0.1$) in mortality among the larvae reared on chicken feed, garden waste, and manures. The effect of cattle and horse manure on mortality was not different, but that of chicken feed and garden waste was comparable. Although the number of live *A. domesticus* larvae was significantly lower ($\alpha = 0.05$) than that of the two species belonging to *Tenebrionidae*, there were no significant differences between *T. molitor* and *Z. morio*. The number of live larvae of all three species significantly decreased with time, and the results of the variance analysis revealed a significant difference in the mortality rate among the species. The effect of the substrates on the mass of live larvae was significantly different ($\alpha = 0.1$) among the larvae reared on chicken feed, green wastes, and manures. The effects of vegetable waste and garden waste were similar to those of cattle and horse manure. After 45 days, the mass of live *A. domesticus* larvae was not significantly different from that of *T. molitor* when fed vegetable and garden waste, but it was significantly lower than that of *T. molitor* when fed the manure substrates. The net weight of *A. domesticus* significantly increased between days 30 and 45 only in larvae fed chicken feed and vegetable waste. On day 45, the minimum weight of live *A. domesticus* larvae reared on cattle manure was 18.92 ± 0.60 g, representing 39.0% of larvae fed chicken feed (48.47 ± 0.77 g). The maximum mass of live larvae was produced by *Z. morio* reared on chicken feed (81.44 ± 1.69 g); however, with only 2.76% mortality, this species produced a live larval mass of 63.65 ± 0.39 g and 63.75 ± 0.77 g when reared on cattle and horse manure, respectively.

Discussion

Nutritional Value of the Larvae

With the hypothesis that diet has a significant effect on the macronutrient composition of larvae, the protein or fat concentrations in the diet for a given species can be tailored. It is necessary to

test diets that represent a wide range of in nutrient concentrations. In this study, chicken feed had the highest protein and carbohydrate concentrations, whereas vegetable waste had the lowest protein concentration with a low carbohydrate concentration, with a difference in means of 86.9% and 84.7%, respectively. The carbohydrate concentration in the waste was not considerably different, whereas the fat concentration in the horse manure diet was 2.38 times higher than that in the vegetable waste. *Acheta domesticus* was rich in proteins and fibre, and poor in fat, whereas *T. molitor* and *Z. morio* were rich in fat and relatively poor in proteins. The different species showed significantly different nutritional composition when reared on different diets, and all low-nutrient value substrates resulted in reduced protein concentrations and increased fat concentration in all three species.

For *T. molitor* and *Z. morio*, BROEKHOVEN et al. (2014) found that the larval protein concentration was relatively stable in diets that differed 2–3-fold in protein concentration and that dietary fat has a significant effect on larval fat concentration. In the present study, the variation in the protein and fat concentrations was low despite the considerable differences in the dietary compositions. Furthermore, the differences between the protein and fat concentrations were lower than the findings of ADÁMKOVÁ et al. (2017b) in the last and penultimate instar stages for both species. They found the protein concentration in *T. molitor* larvae purchased from insect farms (assuming optimal rearing conditions) was 52% (calculated with a CF of 6.25), with a low fat concentration of 31%, whereas the concentration of protein and fat in *Z. morio* was 46% and 35%, respectively. In the case of *T. molitor*, GONZÁLEZ et al. (2019) also reported 48.8% protein and 30.7% fat concentrations. ADÁMKOVÁ et al. (2017a) conducted experiments at 17, 23, and 28 °C, and observed the maximum fat concentration at 23 °C in the last and penultimate instar stages. In *Z. morio* larvae reared on wheat, corn, soybean meal, water, fruits, and vegetables, ARAUJO et al. (2019) reported 46.8% protein and 43.3% lipid concentrations, which are similar to the concentrations recorded in larvae reared on chicken feed in this study. In the case of *A. domesticus* larvae, RUMPOLD and SCHLÜTER (2013) reported 71% protein and 18% fat concentrations, which were obtained from a commercial supplier. This was the closest to our findings when the larvae were reared on chicken feed. Despite the wide range of macronutrient concentrations in the substrates, the nutritional composition of *A. domesticus*, *T. molitor*, and *Z. morio* was species-specific. Overall, to produce larvae with a relatively high fat concentration, *T. molitor* and *Z. morio* are good candidates, whereas *A. domesticus* is predominantly a protein source. However, in agreement with the findings of OONINCX et al. (2015), by choosing an appropriate rearing material, the composition can, to a certain extent, be tailored.

Mass of Live Larvae during Rearing

When MIECH et al. (2016) tested the performance of Cambodian field crickets (*Teleogryllus testaceus*), they found that the larval survival rates on chicken feed, cereals, and green biomass of different plants were similar to those on mono diets, except that certain weeds resulted in lower weights. VEENENBOS and OONINCX (2017) did not find any advantage in feeding additional carrots to improve the survival of *A. domesticus*. A 40% decrease in the survival rate of *A. domesticus* just before adulthood was recorded by VAGA et al. (2020) in a control substrate. In this study, the survival of *A. domesticus* larvae was influenced by the diet. The survival rate was similar to the findings of SORJONEN et al. (2019) for insects reared on alternative diets at the age of 15–45 days, and similarly, low-protein alternative diets resulted in higher mortality. Contrary to our findings, COLLAVO et al. (2005) recorded an almost linear decrease in survival with time at the age of 1–81 days, with a survival rate of 47.5% on human refuse waste, as an example. The effect of temperature lower than the optimal range was presumably low, based on the findings of LACHENICHT et al. (2010). OONINCX et al. (2015) found that the survival of *T. molitor* on low-protein diets (12.9–14.4%), compared with that on

high-protein diets (21.9–22.9%), was low. In this study, *T. molitor* and *Z. morio* did not show a considerable increase in mortality when reared on low-protein diets. The growth rate, biomass accumulation, and population viability were strongly determined by food substrate composition in *A. domesticus*, and the N concentration explained 68% of the variation across treatments, whereas the ratio of N-to-acid detergent fibre explained another 28% of the overall treatment variability (BORDEREAU – ANDERSEN, 1978). The mass of *Z. morio* and *A. domesticus* increased more than that of *T. molitor*, although the weight of the same number of *A. domesticus* larvae was significantly lower than that of *T. molitor* on day 1 of the experiment. The final net mass was still comparable despite the increase in mortality of *A. domesticus* over time. The mass of live larvae is determined by the mortality and the weight of the individual larvae, and their ratio gives the absolute order of the candidates. In this study, *Z. morio* reared on low-value waste showed the highest performance.

Conclusions

Our results confirmed that a high percentage of mixed vegetable waste, garden waste with green biomass of several species, or cattle manure and horse manure could not be considered an optimal rearing substrate to grow *A. domesticus*, *T. molitor*, and *Z. morio* larvae. All low-nutrient value substrates decreased the protein concentration and increased the fat concentration in all three species. Despite the wide range of macronutrient concentrations in the substrates, the nutritional composition of *A. domesticus*, *T. molitor*, and *Z. morio* was species-specific, although the nutritional value of the larvae from the three species was significantly affected by the composition of the rearing substrate. We identified protein and fat combinations for the three species that can be suitable for the feeding programs of animals. In the future, we will evaluate means to prolong the survivability of the larvae at 45 days and relate this to the possible frequency of substrate provision.

References

- ADÁMKOVÁ, A. – ADÁMEK, M. – MLČEK, J. – BORKOVCOVÁ, M. – BEDNÁŘOVÁ, M. – KOUŘIMSKÁ, L. – SKÁCEL, J. – VÍTOVÁ, E. (2017a): Welfare of the mealworm (*Tenebrio molitor*) breeding with regard to nutrition value and food safety. *Potravin. Slovak J. Food Sci.*, 11 (1), 460–465. DOI: <https://doi.org/10.5219/779>
- ADÁMKOVÁ, A. – MLČEK, J. – KOURIMSKÁ, L. – BORKOVCOVÁ, M. – BUSINA, T. – ADÁMEK, M. – BEDNÁROVÁ, M. – KRAJSA, J. (2017b): Nutritional potential of selected insect species reared on the island of Sumatra. *Int. J. Environ. Res. Public Health*, 14 (5), 521. DOI: <https://doi.org/10.3390/ijerph14050521>
- ARAUJO, R.R.S. – DOS SANTOS BENFICA, T.A.R. – FERRAZ, V.B. – SANTOS, E.M. (2019): Nutritional composition of insects *Gryllus assimilis* and *Zophobas morio*: Potential foods harvested in Brazil. *J. Food Compos. Anal.*, 76, 22–26. DOI: <https://doi.org/10.1016/j.jfca.2018.11.005>
- BENZERTIHA, A. – KIERONCZYK, B. – KOŁODZIEJSKI, P. – PRUSZYNSKA-OSZMAŁEK, E. – RAWSKI, M. – JÓZEFIAK, D. – JÓZEFIAK, A. (2020): *Tenebrio molitor* and *Zophobas morio* full-fat meals as functional feed additives affect broiler chickens' growth performance and immune system traits. *Poult. Sci.*, 99 (1), 196–206. DOI: <https://doi.org/10.3382/ps/pez450>

- BOOTH, D.T. – KIDDELL, K. (2007): Temperature and energetics of development in the house cricket (*Acheta domesticus*). *J. Insect Physiol.*, 53 (9), 950–953. DOI: <https://doi.org/10.1016/j.jinsphys.2007.03.009>
- BORDEREAU, C. – ANDERSEN, S.O. (1978): Structural cuticular proteins in termite queens. *Comp. Biochem. Physiol. B Comp. Biochem.* 60 (3), 251–256. DOI: [https://doi.org/10.1016/0305-0491\(78\)90096-2](https://doi.org/10.1016/0305-0491(78)90096-2)
- CADINU, L.A. – BARRA, P. – TORRE, F. – DELOGU, F. – MADAU, F.A. (2020): Insect rearing: Potential, challenges, and circularity. *Sustainability*, 12 (11), 4567. DOI: <https://doi.org/10.3390/su12114567>
- COLLAVO, A. – GLEW, R.H. – HUANG, Y.-S. – CHUANG, L.T. – BOSSE, R. – PAOLETTI, M.G. (2005): House cricket small-scale farming. 515–540. In Paoletti, M.G. (Ed.) *Ecological Implications of Mini Livestock: Potential of Insects, Rodents, Frogs, and Snails*. CRC Press, Boca Raton, 662p. DOI: <https://doi.org/10.1201/9781482294439>
- DUBOIS, M. – GILLES, K.A. – HAMILTON, J.K. – REBERS, P.T. – SMITH, F. (1956): Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 28 (3), 350–356. DOI: <https://doi.org/10.1021/ac60111a017>
- GONZÁLEZ, C.M. – GARZÓN, R. – ROSELL, C.M. (2019): Insects as ingredients for bakery goods. A comparison study of *H. illucens*, *A. domestica* and *T. molitor* flours. *Innov. Food Sci. Emerg. Technol.*, 51, 205–210. DOI: <https://doi.org/10.1016/j.ifset.2018.03.021>
- JANSSEN, R.H. – VINCKEN, J.P. – VAN DEN BROEK, L.A.M. – FOGLIANO, V. – LAKEMOND, C.M.M. (2017): Nitrogen-to-protein conversion factors for three edible insects: *Tenebrio molitor*, *Alphitobius diaperinus*, and *Hermetia illucens*. *J. Agric. Food Chem.*, 65 (11), 2275–2278. DOI: <https://doi.org/10.1021/acs.jafc.7b00471>
- KIM, S.Y. – CHUNG, T.H. – KIM S.H. – SONG, S. – KIM, N. (2014): Recycling agricultural wastes as feed for mealworm (*Tenebrio molitor*). *Korean J. Appl. Entomol.*, 53 (4), 367–373. DOI: <https://doi.org/10.5656/KSAE.2014.10.0.043>
- LACHENICHT, M.W. – CLUSELLA-TRULLAS, S. – BOARDMAN, L. – LE ROUX, C. – TERBLANCHE, J.S. (2010): Effects of acclimation temperature on thermal tolerance, locomotion performance and respiratory metabolism in *Acheta domesticus* L. (Orthoptera: Gryllidae). *J. Insect Physiol.*, 56 (7), 822–830. DOI: <https://doi.org/10.1016/j.jinsphys.2010.02.010>
- LÄHTEENMÄKI-UUTELA, A. – GRMELOVÁ, N. (2016): European law on insects in food and feed. *Eur. Food Feed Law Rev.*, 11 (1), 2–8.
- LUNDY, M.E. – PARRELLA, M.P. (2015): Crickets are not a free lunch: Protein capture from scalable organic side-streams via high-density populations of *Acheta domesticus*. *PLoS ONE*, 10, e0118785. DOI: <https://doi.org/10.1371/journal.pone.0118785>
- MIECH, P. – BERGGEN, A. – LINDBERG, J.E. – CHHAY, T. – KHIEU, B. – JANSSON, A. (2016): Growth and survival of reared Cambodian field crickets (*Releogryllus testaceus*) fed weeds, agricultural and food industry by-products. *J. Insects Food Feed*, 2 (4), 285–292. DOI: <https://doi.org/10.3920/JIFF2016.0028>
- MORALES-RAMOS, J.A. – ROJAS, M.G. – DOSSEY, A.T. (2018): Age-dependent food utilisation of *Acheta domesticus* (Orthoptera: Gryllidae) in small groups at two temperatures. *J. Insects Food Feed*, 4 (1), 51–60. DOI: <https://doi.org/10.3920/JIFF2017.0062>
- OONINCX, D.G.A.B. – van BROEKHOVEN, S. – van HUIS, A. – van LOON, J.J.A. (2015): Feed conversion, survival and development, and composition of four insect species on diets

composed of food by-products. *PLoS ONE*, 10, e0144601. DOI: <https://doi.org/10.1371/journal.pone.0144601>

ORTIZ, J.A.C. – RUIZ, A.T. – MORALES-RAMOS, J.A. – THOMAS, M. – ROJAS, M.G. – TOMBERLIN, J.K. – YI, L. – HAN, R. – GIROUD, L. – JULLIEN, R.L. (2016): Insect mass production technologies. *Insects as Sustainable Food Ingredients: Production, Processing, and Food Applications*; 153–201. DOI: <https://doi.org/10.1016/B978-0-12-802856-8.00006-5>

RUMBOS, C.I. – KARAPANAGIOTIDIS, I.T. – MENTE, E. – PSOFAKIS, P. – ATHANASSIOU, C.G. (2020): Evaluation of various commodities for the development of the yellow mealworm, *Tenebrio molitor*. *Sci. Rep.*, 10, 11224. DOI: <https://doi.org/10.1038/s41598-020-67363-1>

RUMPOLD, B.A. – SCHLÜTER, O.K. (2013): Nutritional composition and safety aspects of edible insects. *Mol. Nutr. Food Res.*, 57 (5), 802–823. DOI: <https://doi.org/10.1002/mnfr.201200735>

SORJONEN, J.M. – VALTONEN, A. – HIRVISALO, E. – KARHAPÄÄ, M. – LEHTOVAARA, V.J. – LINDGREN, J. – MARNILA, P. – MOONEY, P. – MÄKI, M. – SILJANDER-RASI, H. – TAPIO, M. – TUISKULA-HAAVISTO, M. – ROININEN, H. (2019): The plant-based by-product diets for the mass-rearing of *Acheta domesticus* and *Gryllus bimaculatus*. *PLoS ONE*, 14, e0218830. DOI: <https://doi.org/10.1371/journal.pone.0218830>

VAGA, M. – BERGGREN, A. – PAULY, T. – JANSSON, A. (2020): Effect of red clover-only diets on house crickets (*Acheta domesticus*) growth and survival. *J. Insects Food Feed*, 6 (2), 179–189. DOI: <https://doi.org/10.3920/JIFF2019.0038>

VARELAS, V. (2019): Food wastes as a potential new source for edible insect mass production for food and feed: A review. *Fermentation*, 5 (3), 81. DOI: <https://doi.org/10.3390/fermentation5030081>

VARELAS, V. – LANGTON, M. (2017): Forest biomass waste as a potential innovative source for rearing edible insects for food and feed – A review. *Innov. Food Sci. Emerg. Technol.* 41, 193–205. DOI: <https://doi.org/10.1016/j.ifset.2017.03.007>

VEENENBOS, M.E. – OONINCX, D.G.A.B. (2017): Carrot supplementation does not affect house cricket performance (*Acheta domesticus*). *J. Insects Food Feed* 3, 217–221. DOI: <https://doi.org/10.3920/JIFF2017.0006>

Authors

Richárd PINTÉR

PhD student

Institute of Food Science and Technology, Center for Food Technology, Department of Animal Product and Food Preservation Technology, 1118-Budapest Ménesi st. 43-45., H-1118 Budapest, Hungary
pinterichard@gmail.com

György FEKETE

levelező szerző

PhD student

Institute of Environmental Sciences, Hungarian University of Agriculture and Life Sciences, Páter Károly 1, H-2100 Gödöllő, Hungary
fekete.gyorgy@uni-mate.hu

Zsolt István VARGA

PhD student

Institute of Environmental Sciences, Hungarian University of Agriculture and Life Sciences,
Páter Károly 1, H-2100 Gödöllő, Hungary
varga.zsolt.istvan@uni-mate.hu

Dr. Csaba GYURICZA DSc

professor, rector

Institute of Crop Production, Hungarian University of Agriculture and Life Sciences,
Páter Károly 1, H-2100 Gödöllő, Hungary
gyuricza.csaba@uni-mate.hu

Dr. László ALEKSZA PhD

associate professor

Institute of Environmental Sciences, Hungarian University of Agriculture and Life Sciences,
Páter Károly 1, H-2100 Gödöllő, Hungary
aleksza.laszlo@uni-mate.hu

A műre a Creative Commons 4.0 standard licenc alábbi típusa vonatkozik: [CC-BY-NC-ND-4.0.](https://creativecommons.org/licenses/by-nc-nd/4.0/)

