

RESURGENCE OF *TOMATO SPOTTED WILT VIRUS* ON PEPPER (*CAPSICUM ANNUUM* L.) PLANTS IN SZENTES REGION

**István Tóbiás^{1*}, Gábor Csilléry², Asztéria Almási¹,
Katalin Salánki¹, Zsófia Csömör^{1,3}, László Palkovics³**

¹*MTA ATK Növényvédelmi Intézet, Budapest*

²*Budakert Kft., Budapest*

³*Budapesti Corvinus Egyetem, Kertészettudományi Kar, Budapest*

*tobias.istvan@agrar.mta.hu

Abstract

In Hungary *Tomato spotted wilt virus* was considered as an important pathogen since the mid-nineties. The introduction and spread of western flower thrips (*Frankliniella occidentalis*), an efficient TSWV vector, in that time certainly played an important role in TSWV emergence. Management of TSWV control was first directed against the thrips and weeds. Later on *Tsw* resistant gene was introduced into different types of pepper. In 2012 heavy crop losses were observed on TSWV resistant pepper varieties in Szentes region. Systemic virus symptoms on leaves and fruits and decline were observed in TSWV resistant cultivars, caused by resistance breaking strain of *Tomato spotted wilt virus*.

Key-words: *Tomato spotted wilt virus*, pepper, resistance, resistance breaking strain

Összefoglalás

Hazánkban a paradicsom foltos hervadás vírus (*Tomato spotted wilt virus*, TSWV) az 1990-es évek közepén vált jelentős kórokozóvá. Ebben döntő szerepet játszott a vírus hatékony vektorának, a nyugati virág tripsznek (*Frankliniella occidentalis*) a Magyarországra történő behurcolása. A betegség elleni védekezés eleinte a vektor ellen irányult, majd később a nemesítő intézetek TSWV rezisztens fajtákat állítottak elő szinte minden fajtatípusból, a Tsw rezisztenciagén sikeres beépítésével. 2012-ben Szentés környékén nagymértékű fertőzést figyeltünk meg a TSWV rezisztens paprikafajtákon. A jellegzetes levél és bogyó tünetek mellett nagyarányú növénypusztulást is észleltünk. A fertőzést a paradicsom foltos hervadás vírus rezisztencia áttörő törzse okozta.

Kulcsszavak: paradicsom foltos hervadás vírus, paprika, rezisztencia, rezisztencia áttörő törzs

Introduction

Tomato spotted wilt virus (TSWV) is the type member of the genus *Tospovirus* (family *Bunyaviridae*), causes an important disease of horticultural and agronomic crops. The virus distributed worldwide is having extremely broad host range and is now considered as one of the ten most economically destructive plant viruses (Adkins 2000, Moyer 1999, Tomlinson 1987). TSWV is transmitted by thrips in a persistent manner, only the larvae can acquire the virus, which multiplies in the vector and the adults can transmit (Ullmann et al 1992). The virion varies in size from 80 to 120 nm and has spherical enveloped character (Prins and Goldbach 1998). The genome of TSWV consists of three ssRNA segments:

small (S) and medium (M) RNAs have ambisense coding strategies, whereas the large (L) RNA is of negative polarity (Fig. 1.).

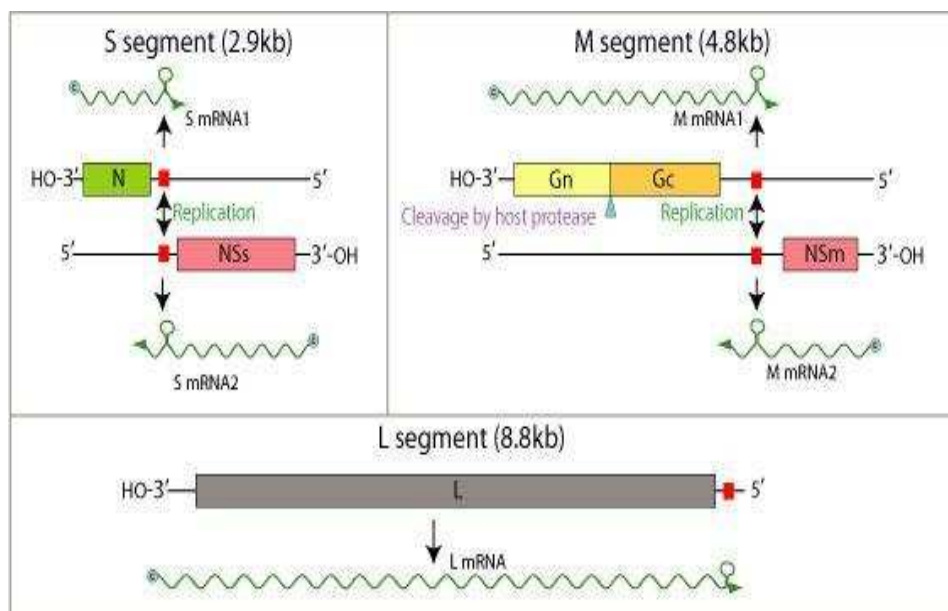


Figure 1. *Tomato spotted wilt virus genome*

In Hungary TSWV was described in 1972 (Ligeti and Nagy 1972), but the virus was not considered as an important pathogen. In 1995 very severe damage of TSWV infection was observed in tomato and pepper production in the Szentes vegetable growing region. The introduction and spread of western flower thrips (*Frankliniella occidentalis*), an efficient TSWV vector, in that time certainly played an important role in TSWV emergence (Csilléry et al 1995, Gáborjányi et al 1995, Jenser and Tusnádi 1989, Jenser 1995).

Management of TSWV control was first directed against the thrips using different insecticides or plastic traps, and against weeds as host plants of the virus and the thrips. Later on *Tsw* resistant gene from *Capsicum chinense* PI-

152225 és PI-159236 (Black et al 1996) was introduced into different types of pepper (conical white, long pale green hot and sweet, tomato shape, spice pepper and blocky types) (Csilléry unpublished). Pepper cultivars carrying *Tsw* resistance gene upon TSWV inoculation show necrotic local lesions on the leaves or other parts of the plant without systemic infection.

It was demonstrated that TSWV can adapt very rapidly to plant resistance, and the *Tsw* resistance gene was broken down only a few years after its deployment in pepper crops (Roggero et al 2002, Thomas-Carroll and Jones 2003, Margaria et al 2004, Sharman and Persey 2006).

In 2010 and 2011 sporadically, but in 2012 and 2013 more frequently systemic virus symptoms were observed on resistant pepper cultivars in Szentes region (Bese et al. 2012, Csilléry et al 2012, Salamon et al 2010). The presence of new resistance breaking strain of TSWV was proved by virological (test-plant, serological and RT-PCR) methods.

Materials and Methods

Virus isolates. TSWV isolates originated from pepper cultivars susceptible and resistant

against TSWV from Szentes region. Fruit samples were collected from plants exhibiting typical symptoms of virus infection such as stunting, mosaic, chlorotic and/or necrotic spots, rings and distortion on the leaves and fruits (Fig. 2). The isolates were investigated on test plants, ELISA serological tests, RT-PCR and maintained by mechanical inoculation on *Nicotiana tabacum* cv. *Xanthi-nc* plants. The original samples were kept at $-70\text{ }^{\circ}\text{C}$.

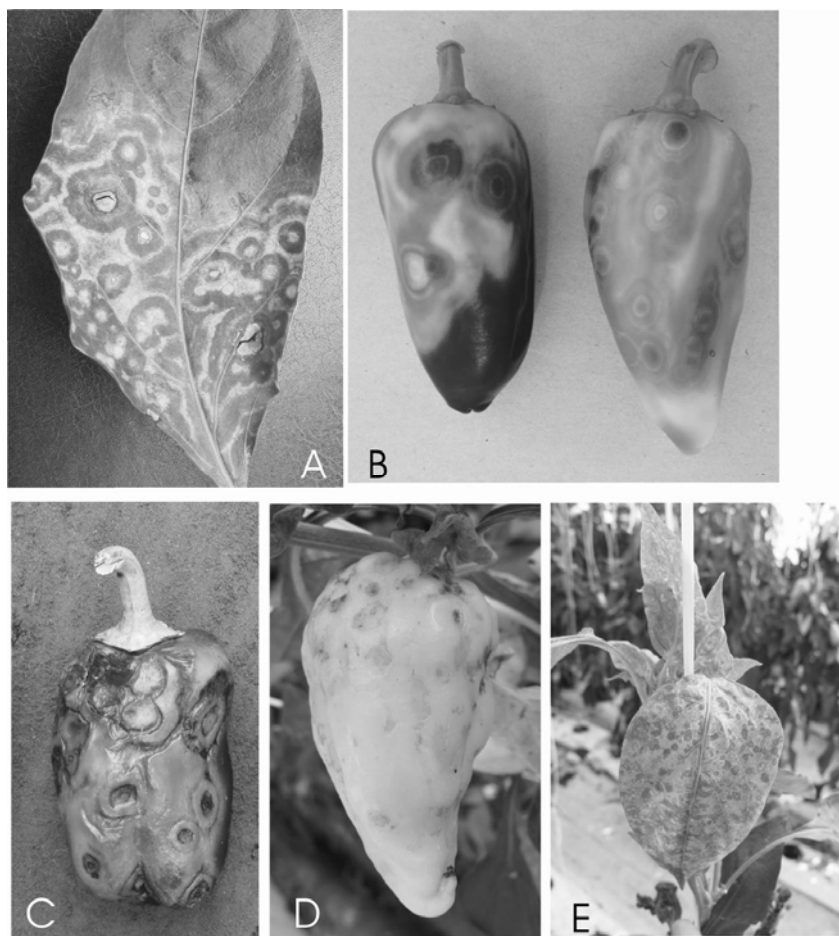


Figure 2. Symptoms of TSWV infection on leaves (A) and fruits (B) on susceptible cultivars, and on fruits of resistant cultivar (C). Systemic symptoms of resistance breaking TSWV isolate on resistant cultivars (D and E)

Results

The collected samples showed typical symptoms of *Tomato spotted wilt virus* infection. The virus was transmitted by mechanical inoculation onto test

plants. On *Nicotiana tabacum* cv. *Xanthi-nc* plants chlorotic and necrotic spots and rings on inoculated leaves and systemic mosaic or necrotic rings or necrosis were observed (Fig. 3). Slight differences on symptoms were observed among different isolates independently whether originated from TSWV susceptible or resistant pepper cultivars

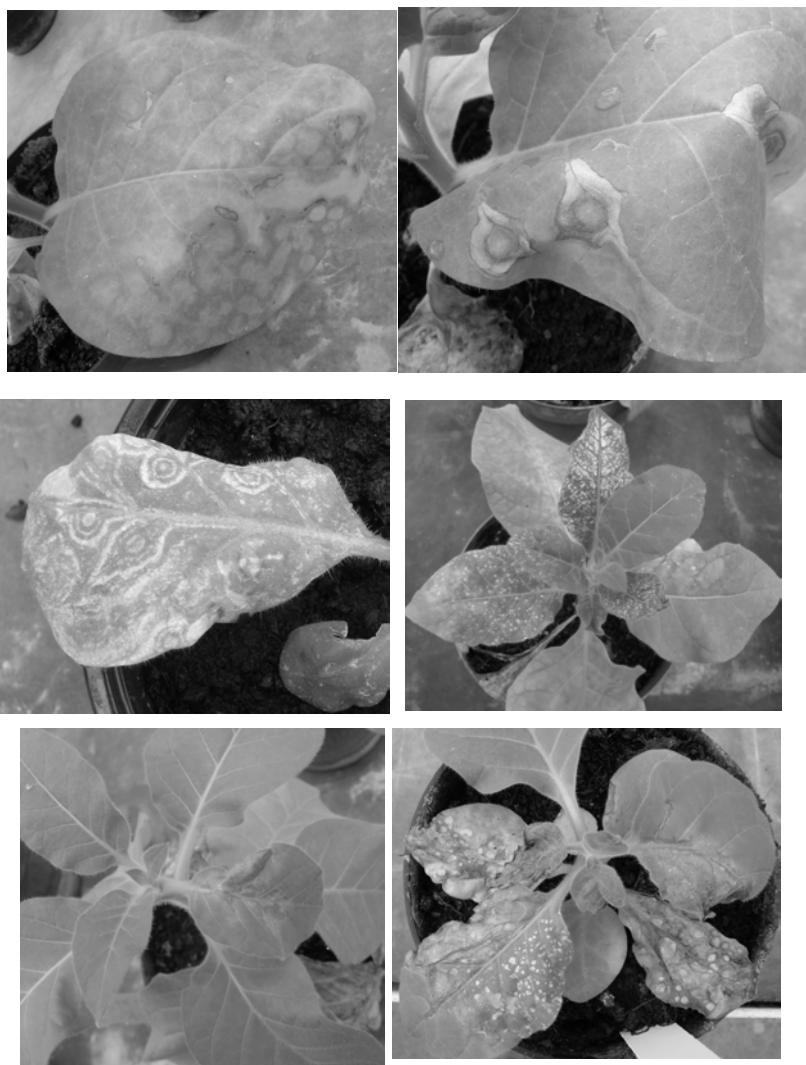


Figure 3. Different type of local and systemic symptoms on Nicotiana tabacum cv. Xanthi-nc plant caused by different TSWV isolates

Some TSWV isolates were inoculated onto *Emilia sonchifolia* (this plants are systemically infected by TSWV and generally used to maintained the TSWV isolates) and only local necrotic symptoms were observed in contrast with other TSWV isolates (Fig. 4).



Figure 4. Local symptoms of Emilia sonchifolia after inoculation of Tomato spotted wilt virus isolated in Hungary.

TSWV specific PCR-product was amplified by RT-PCR method (Fig. 5). Our results confirmed the presence of *Tomato spotted wilt virus* both in TSWV susceptible and resistance cultivars in Hungary. Our results confirmed the presence of the resistance breaking isolate of *Tomato spotted wilt virus* in Hungary. Further investigations needed to characterize the resistance breaking TSWV isolates from Hungary.

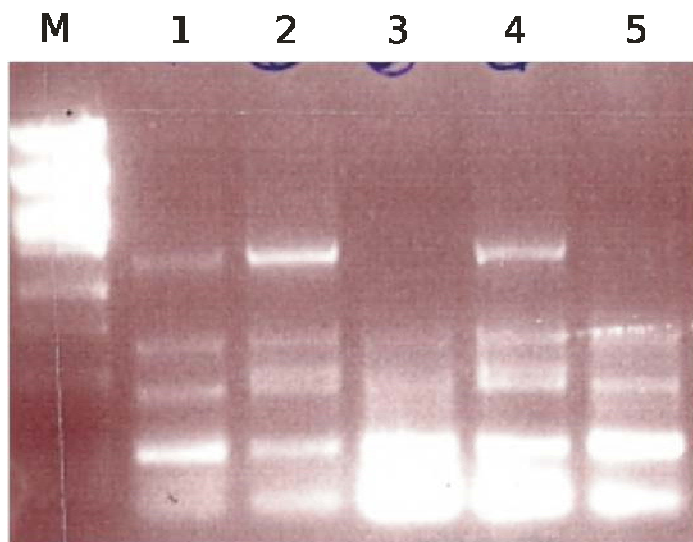


Figure 5. Separation of amplified RT-PCR products of TSWV infected pepper plants on 1 % agarose gel stained with ethidium bromide. M– DNA lenght marker Pst I digested λ DNA, Lane 1, 2 and 4 TSWV infected pepper, Lane 3 uninfected and Lane 5 healthy pepper plants.

Summarizing our investigations on TSWV resurgence in Hungary we can conclude that the first TSWV epidemic in mid 1990 years was connected with the introduction and spread of western flower thrips (*Frankliniella occidentalis*), an efficient TSWV vector, while the second emergence in 2010 and later was due to the appearance of resistance breaking isolates of TSWV.

Acknowledgement

Present article was published in the frame of the project TÁMOP-4.2.2.A-11/1/KONV-2012-0064. „Regional effects of weather extremes resulting from climate change and potential mitigation measures in the coming decades.” The project is realized with the support of the European Union, with the co-funding of the European Social Fund.

References

Adkins S. 2000. Tomato spotted wilt virus – positive steps towards negative succes. *Molecular Plant Pathology* 1, 151-157.

Bese G., Krizbai L., Takács A.P. 2012. A paradicsom foltos hervadás vírus (*Tomato spotted wilt virus*, TSWV) rezisztencia áttörő törzs első megjelenése Magyarországon. *Növényvédelmi Tudományos Napok*, Budapest pp 49.

Boiteaux, L. S., and Nagata, T. 1993. Susceptibility of *Capsicum chinense* PI159236 to *Tomato spotted wilt virus* isolates in Brazil. *Plant Dis.* 77:219.

Csilléry G., Gáborjányi R., Tóbiás I. és Jenser G. 1995. Új paprika és paradicsom kórokozó. Paradicsom foltos hervadás vírus. *Kertészet és Szőlészet*, 29:8-9.

Gáborjányi R., Csilléry G., Tóbiás I., Jenser G. 1995. *Tomato spotted wilt virus: A new threat for pepper production in Hungary*. IXth Eucapia Meeting, Budapest, 159-160.

German T. L., Adkins S., Witherell A., Richmond K. E., Knaack W. R., Willis D. K. (1995) Infection of *Arabidopsis Thaliana* Ecotype Columbia by *tomato spotted wilt virus*. *Plant Mol Biol Rep* 13(2):110–117

Jahn, M., Paran, I., Hoffmann, K., Radwanski, E. R., Livingstone, K. D., Grube, R. C., After- goot, E., Lapidot, M., and Moyer, J. W. 2000. Genetic mapping of the *Tsw* locus for resistance to the *Tospovirus* *Tomato spotted wilt virus* in *Capsicum* spp. and its relationship to the *Sw-5* gene for resistance to the same pathogen in tomato. *Mol. Plant-Microbe In- teract.* 13:673-682.

Jenser G and Tusnádi Cs. K. 1989: A nyugati virágtripsz (*Frankliniella occidentalis* Pergande) megjelenése Magyarországon. (The appearance of

Frankliniella occidentalis Pergande in Hungary). Növényvédelem 25, 389–392.

Jenser G. 1995: A tripszek szerepe a paradicsom bronzfoltosság vírus terjedésében (The role of the *Thysanoptera* species in the spread of *tomato spotted wilt tospovirus*). Növényvédelem 31, 541–545.

Ligeti L and Nagy Gy. 1972: A *Lycopersicum* vírus 3 dohányültetvényeink új, veszedelmes kórokozója. Dohányipar: 41–43.

Margaria, P., Ciuffo, M., and Turina, M. 2004. Resistance breaking strains of *Tomato spotted wilt virus (Tospovirus-Bunyaviridae)* on resistant pepper cultivars in Almeria (Spain). Plant Pathol. 53:795.

Moyer, J.W., 1999. *Tospoviruses (Bunyaviridae)*. In: Granoff, A., Webster, R.G. (Eds.), Encyclopedia of Virology. Academic Press, London, pp. 1803/1807.

Prins M., Goldbach R. 1998. The emerging problem of tospovirus infection and nonconventional methods of control. Trends Microbiol. 6, 31–35.

Roggero, P., Melani, V., Ciuffo, M., Tavella, L., Tedeschi, R., and Stravato, V. M. 1999. Two field isolates of *Tomato spotted wilt Tospovirus* overcome the hypersensitive response of a pepper (*Capsicum annuum*) hybrid with resistance introgressed from *C. chinense* “PI152225”. Plant Dis. 83:965.

Roggero, P., Masenga, V., and Tavella, L. 2002. Field isolates of *Tomato spotted wilt virus* overcoming resistance in pepper and their spread to other hosts in Italy. Plant Dis. 86:950–954.

Salamon P., Nemes K., Salánki K. 2010. A paradicsom foltos hervadás vírus (*Tomato spotted wilt virus*, TSWV) rezisztenciatoró törzsének első izolálása paprikáról (*Capsicum annuum* L) Magyarországon. Növényvédelmi Tudományos Napok pp 23.

Sharman M., Persley D.M. 2006. Field isolates of Tomato spotted wilt virus overcoming resistance in *Capsicum* in Australia. *Australasian Plant Pathology* 35, 123-128.

Thomas-Carrol, M. L., and Jones, R. A. C. 2003. Selection, biological properties and fitness of resistance-breaking strains of *Tomato spotted wilt virus* in pepper. *Ann. Appl. Biol.* 142:235-243.

Ullman, D.E., Cho, J.J., Mau, R.F.L., Westcot, D.M., Cantone, D.M., 1992. Midgut epithelial cells act as a barrier to *Tomato spotted wilt virus* acquisition by adult western flower thrips. *Phytopathology* 85, 456/463

