

INVESTIGATION OF THE CAUSATIVE AGENT OF A VIRUS-LIKE SYMPTOM IN GRAPEVINE

Dana Khrais^{1,2}, Emese Demián^{1,2}, Éva Várallyay¹*

*¹NARIC, Agricultural Biotechnology Institute, Molecular Plant Pathology
Group*

H-2100 Gödöllő, Szent-Györgyi A. str. 4.

*²Hungarian University of Agriculture and Life Sciences, H-2100, Gödöllő, Páter
Károly str.*

** Corresponding author, demian.emese@abc.naik.hu*

Abstract

Our group has been carrying on virus diagnostic surveys since 2013 using a special method: small RNA HTS. In 2018 strange symptoms, suggesting the presence of grapevine red blotch-associated virus (GRBaV), appeared at a Pinot Noir plantation. The virus was described in California in 2013 and its presence, except two descriptions (India and South-Korea) is confined to the North-American continent. To reveal the causative agent of the appeared symptoms we purified RNA from the symptomatic grapevine and made virus diagnostics by small RNA HTS. We confirmed the results using virus specific RT-PCR. Our results show that not the presence of the assumed virus cause the observed symptoms, moreover GRBaV is not present at the investigated vineyard. However, the plantation was infected with several other viruses. We

think that coexistence of several different viruses together with the inhomogeneity in the soil can both contribute in the symptom development, however to clarify this question further investigations would be needed.

Keywords: grapevine red blotch-associated virus, GLRaV-1, small RNA HTS, RT-PCR

Összefoglalás

Kutatócsoportunk 2013 óta végez vírusdiagnosztikai felméréseket szőlőültetvényekben egy új módszer, a kis RNS HTS használatával. 2018-ban egy Pinot noir ültetvényen furcsa, a szőlő vörös foltosodás vírus (GRBaV) jelenlétére utaló tüneteket figyeltek meg. A vírust 2013-ban Kaliforniában írták le, elterjedése az EPPO nyilvántartása szerint 2 kivételtől eltekintve (India és Dél-Korea) az észak-amerikai földrészre korlátozódik. A tünetek okainak felderítésére a jellemző tüneteket mutató szőlők RNS-éből kis RNS szekvenálással végeztünk vírusdiagnosztikát. A kapott eredményeket egy független módszerrel, RT-PCR-rel igazoltuk. Eredményeink azt mutatják, hogy a sajátos tünetek kialakulásában nem a feltételezett vírus játszott szerepet, a GRBaV nincs jelen a vizsgált ültetvényen. Az ültetvény viszont több vírussal is fertőzött volt. A tünetek kialakulásának az oka ez esetben a különböző vírusok együttes fertőzésében, az ültetvény talajának inhomogenitásában keresendő, de pontos megállapításához további vizsgálatok szükségesek.

Kulcsszavak: szőlő vörös foltosodás vírus, GLRaV-1, kis RNS HTS, RT-PCR

Introduction

Grapevine is considered as one of the major fruit crops in the world based on hectares cultivated and economic value. It can be infected with several (more than 80 is described until now) viruses which presence can affect not only its growth, but the quality of important

characteristics (berry weight and colour and sugar content, etc.). Grapevine red blotch-associated virus (GRBaV) was described in California, from a vineyard showing red blotch disease using HTS of ds RNAs (Rwahnih et al., 2013) and was proved to be the causative agent of the disease later (Yepes et al., 2018). It is a member of the Geminiviridae family, having a single circular DNA genome and its presence was proved to have inferior effect on berry development (Blanco-Ulate et al., 2017). The disease symptoms in red varieties include reddening of regions within leaf blades, along with red veins and petioles and delayed fruit maturity. In white varieties, leaves may develop yellow or chlorotic that is similar to leafroll-diseased vines. Asymptomatic vines can remain productive, but they also harbour viruses and act as potential reservoirs for virus spread to susceptible vines. It is graft transmittable, it could originated from a wooded riparian area by a supposed new vector (Cieniewicz et al., 2017). Grapevine virologists highlight that symptoms can be very similar to leafroll disease, with an exception that in GRBaV infected plants margin of the leaf stay flat and instead of green, pink veins appear (Sudarshana et al., 2015). As small RNA HTS can detect the presence of DNA viruses (Pooggin, 2018), we used this method to reveal the causative agent of the observed, virus-like symptoms.

Materials and Method

At the beginning of June 2018, the above symptoms: red blotches and red veins appeared at a Pinot noir plantation at Somogy (South of the Balaton) (Figure 1/A). At the vineyard plants with leafroll symptoms also appeared (Figure 1/B), and these two markedly different symptoms was altered even within a row.

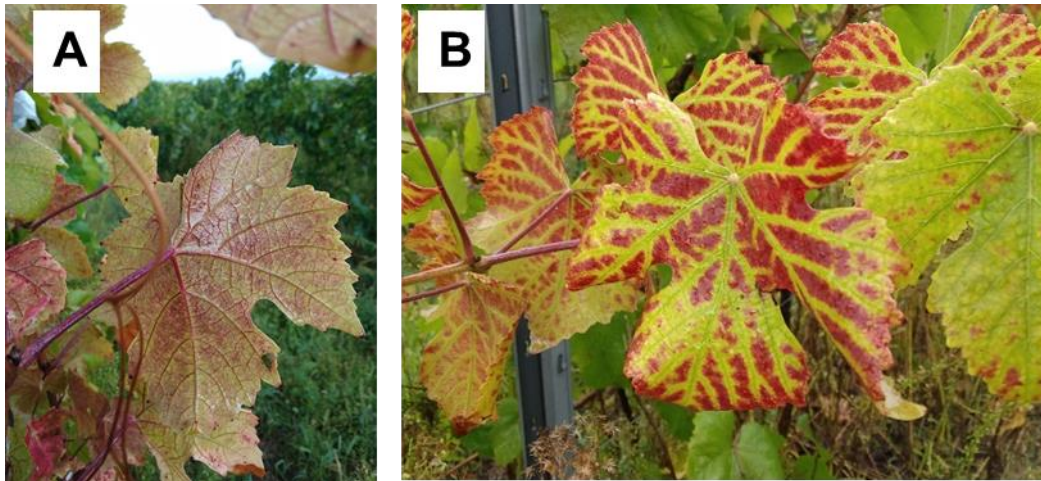


Figure 1. Pictures of the surveyed Pinot noir grapevines showing A/ red blotch-like, B/ leafroll-like symptoms.

Samples were collected from 4 individual plants of the same row, showing either red blotch-like (1, 3) or leafroll-like (2, 4) symptoms. We extracted RNA using a CTAB method and prepared two small RNA sequencing libraries (RB from plant 1 and 3 and LR from plant 2 and 4) according to our adapted protocol (Czotter et al., 2018a).

The sequenced reads were analysed using Qiagen CLC Genomic workbench. The reads were trimmed, both redundant and non-redundant list of sequences were prepared. This later one was used for contig building. Virus diagnostics were done by BLAST search of assembled contigs using all plant hosted viruses in the NCBI. The result list was ordered according to their lowest E-value. The reads (both redundant and non-redundant) were mapped to the GRBaV reference genome, and for other viruses which were present according to the analysis. Based on this analysis the consensus sequences were prepared and the coverage of the viral genome by small RNA reads were calculated. Threshold of virus presence was set to at least 1 virus specific contig, and higher than 60% coverage of the genome.

To validate the results of the small RNA HTS and directly test the presence of GRBaV, RT-PCR was carried out. After cDNA synthesis, an actin test was used to check the cDNA quality. This cDNA was used as a template to validate the presence of the presenting grapevine viruses

by RT-PCR using virus specific primers (for primer sequences please check (Czotter et al., 2018b). We also tested the cDNA with published GRBaV primers (Rwahnih et al., 2013).

Results and discussion

Results of the small RNA HTS show that several viruses: GLRaV-1, GVA, GFkV, GPGV and possibly GVB and viroids: HSVd, GYSVd-1 and 2 were present in the tested plants (Table 1), but neither GRBaV positive contig, nor reads mapped to the GRBaV genome were identified. Moreover, we could not get any product in the RT-PCR reaction using virus specific primers.

Table 1. Summary of the bioinformatics analysis together with the RT-PCR validation. GRBaV: grapevine red blotch-associated virus. Numbers indicate PCR positive samples out of the 2 which served for small RNA library preparation.

	type of analysis	viruses							viroids	
		GRBaV	GLRaV-1	GVA	GVB	GFkV	GRSPaV	GPGV	HSVd	GYSVd-1/2
RB library	small RNA HTS	0	+	+	0	0	0	+	+	+
	RT-PCR	0/2	1/2	1/2	0/2	1/2	2/2	2/2	2/2	2/2
LR library	small RNA HTS	0	+	+	?	+	0	+	+	+
	RT-PCR	No	2/2	2/2	0/2	2/2	2/2	2/2	2/2	2/2

GLRaV-1: grapevine leafroll associated virus-1, GVA: grapevine virus A, GVB: grapevine virus B, GFkV: grapevine fleck virus, GRSPaV: grapevine rupestris stem pitting-associated virus, GPGV: grapevine Pinot gris virus, HSVd: hop stunt viroid, GYSVd-1 and 2: grapevine yellow speckled viroid-1 and 2.

We have found severe infection with GLRaV-1 and also the presence of several other viruses and viroids (Table 1.) In case of GLRaV-1, GVA, GPGV and the viroids our RT-PCR results verified the result of the small RNA HTS (Figure 2). We could not prove the presence of GVB, but we have found only 1 GVB positive contig and less than 40% coverage of the genome, which indicate a false positive hit during the analysis. In RB library GFkV was not detected by

small RNA HTS, but in RT-PCR one of the plants showed infection. RNA from the other plant could dilute the sample what was used for small RNA sequencing, why we failed to detect it by this method. For GRSPaV we found the same contradiction, but this is what we usually experience in case of this virus. There were very few GRSPaV derived small RNA reads in the sample while the virus was present. One explanation of this can be that it was proved that this virus can have a positive effect on the grapevine physiology why defence mechanism against it could be suppressed during the evolution.

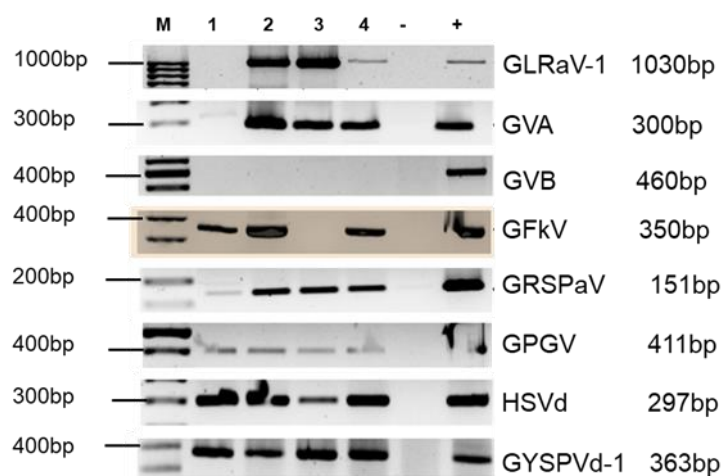


Figure 2. RT-PCR analysis for testing the presence of different viruses in the four plants which small RNA was sequenced.

In summary presence of several different viruses seemed random in the plants, thus we could not correlate any special combination with the appeared symptoms.

Conclusions

Our results showed that although red blotch symptoms appeared, GRBaV was not identified in the investigated Hungarian vineyard. Although it seemed possible for us to detect the presence of several different viruses and viroids, we cannot make any hypothesis about their contribution in the observed symptom development. Distribution of the nutrient in the soil of this vineyard

is very patchy why it is possible that shortage of some of them occurs quite randomly. Combination of these abiotic effects with the virus infections could lead finally or play role in the observed symptom development, but to find out the real causative agent we need further investigations. Moreover, these ambiguous results highlight the importance of the cooperation of classical and molecular virologist to reveal practical importance of the detected virus infections and explain or disclose possible causes of the emerging symptoms.

Acknowledgements

This work was supported by a grant from NKFIH: 119783. Dana Khrais is an MSc student at SZIU. Emese Demian a PhD student of the Doctoral school of Biological Sciences at SZIU.

References

- Blanco-Ulate, B., Hopfer, H., Figueroa-Balderas, R., Ye, Z., Rivero, R. M., Albacete, A., *et al.* 2017. Red blotch disease alters grape berry development and metabolism by interfering with the transcriptional and hormonal regulation of ripening. *Journal of Experimental Botany*. **68**. 1225-1238.
- Cieniewicz, E. J., Pethybridge, S. J., Gorny, A., Madden, L. V., McLane, H., Perry, K. L., *et al.* 2017. Spatiotemporal spread of grapevine red blotch-associated virus in a California vineyard. *Virus Research*. **241**. 156-162.
- Czotter, N., Molnár, J., Pesti, R., Demián, E., Baráth, D., Varga, T., *et al.* 2018a. Use of siRNAs for Diagnosis of Viruses Associated to Woody Plants in Nurseries and Stock Collections. In: *Viral Metagenomics: Methods and Protocols*. (Pantaleo, V. and Chiumenti, M., eds.). New York, NY: Springer New York, pp. 115-130.

Czotter, N., Molnar, J., Szabo, E., Demian, E., Kontra, L., Baksa, I., *et al.* 2018b. NGS of Virus-Derived Small RNAs as a Diagnostic Method Used to Determine Viromes of Hungarian Vineyards. *Frontiers in Microbiology*. **9**. 122.

Pooggin, M.M. 2018. Small RNA-Omics for Plant Virus Identification, Virome Reconstruction, and Antiviral Defense Characterization. *Frontiers in Microbiology*. **9**. 2779.

Rwahnih, M.A., Dave, A., Anderson, M.M., Rowhani, A., Uyemoto, J.K., Sudarshana, M.R. 2013. Association of a DNA Virus with Grapevines Affected by Red Blotch Disease in California. *Phytopathology*. **103**. 1069-1076.

Sudarshana, M.R., Perry, K.L., Fuchs, M.F. 2015. Grapevine Red Blotch-Associated Virus, an Emerging Threat to the Grapevine Industry. *Phytopathology*. **105**. 1026-1032.

Yepes, L.M., Cieniewicz, E., Krenz, B., McLane, H., Thompson, J.R., Perry, K.L., *et al.* 2018. Causative Role of Grapevine Red Blotch Virus in Red Blotch Disease. *Phytopathology*. **108**. 902-909.

Instructions to Authors

The aim of *Georgikon for Agriculture* is to publish original papers in all fields of agriculture and related topics. They may include new scientific results, short communications, critical review articles, conference reviews and letters to the Editor.

Manuscripts should be sent in English **electronically** (farsang.sandorne@uni-mate.hu and anda.angela@uni-mate.hu).

Manuscripts are anonymously reviewed, and if necessary returned to the authors for correction. Proofs should be checked and returned to the Editor within 48 hours after receipt. Publishing in the Journal is free of charge.

The manuscript should be in double spaced typing in justified alignment using Times New Roman fonts, 12 pt character size except for the title, name and affiliation block. The manuscript length should not exceed 16 printed pages including tables and figures. Metric (SI) symbols should be used. Main section names (*Abstract, Összefoglalás, Introduction, Materials and Methods, Results, Discussion, References, Acknowledgement* if applicable, *Tables and Figures*) should be aligned to the centre in italic bold 12 pt size characters. Minor headings are set in italic type, aligned at the left. Leave one blank line between sections.

Title: Should be short, compact and relevant, expressing the contents of the work. The recommended limit is 12 words. Type title of the paper in centred bold capital letters, in 16 pt size characters aligned to the centre of the line.

Author(s) name(s): Leave one blank line before the name- and affiliation block. Please give the whole name of all author(s) and address(es). In the case of two or more authors, the author's names should be followed by numbering in the upper case to separate their addresses. An asterisk (*) follows the corresponding author's name. Provide E-mail address for the correspondent author. Name and affiliation should be typed using centred alignment, italic 14 pt size characters followed by one blank line.

Abstract: The title should be followed by an Abstract, containing the scope of the work and the principal findings in fewer than 200 words. Leave one blank line after the abstract and give maximum 5 to 8 keywords.

Összefoglalás: The keywords should be followed by a summary, written in Hungarian, entitled - Összefoglalás - not longer than 300 words.

Introduction: This part should state briefly the nature and purpose of the work and cite recent important research results in the area. References should be cited as follows: ...as observed by Hatfield and Idso (1997); or in parentheses:were found (Hatfield et al., 1998; Jackson and Hatfield, 1997).

When referring to several papers published in the same year by the same author, the year of publication should be followed by letters a,b,c etc. Cite only essential references.

Materials and methods: should contain the precise description of materials, methods, equipments, experimental procedure and statistical methods used, in sufficient detail.

Results: This part of the paper should present the experimental data clearly and concisely together with the relevant tables and figures.

Discussion: This part should focus on the interpretation of the experimental findings, contain the conclusions drawn from the results, discussing them with respect to the relevant literature.

Acknowledgement: grants and various kinds of assistance may be mentioned here.

References: The list of references should be arranged alphabetically by the authors surnames. Make sure that all references in the paper are listed in this part and vice versa. If necessary cite papers not published yet as 'unpublished data' or 'pers.com.'.

The reference in the case of journal papers should contain: name(s) and initials of all author(s), year of publication, title of article, name of journal, volume number and pages. Use italic letters for the journal name and bold letters for volume number. E.g. Bauer, P.J., Frederick, R.J.,

Bradow, E.J., Sadler, E.J. and Evans, D.E. 2000. Canopy photosynthesis and fiber properties of normal- and late-planted cotton. *Agronomy Journal*. **92**. 518-523.

Reference for books should contain name(s) of author(s), year of publication, title of the book, publisher, place of publication and pages. E.g. Storch, H. von. and Flöser, G. 2000. Models in Environmental Research. Springer-Verlag, Berlin/Heidelberg, 152-158.

Example of a reference for chapter in a proceedings volume: Cagirgan, M.J., and C. Toker. 1996. Path-coefficient analysis for grain yield and related characters under semiarid conditions in barley. p: 607-609. *In* A. Slinkard et al. (ed) Proc. Int. Oat Conf., 5th Int. Barley Genet. Symp., 7th Vol. 2. Univ. of Saskatchewan Ext. Press, Saskatoon, Canada.

Figures: Number the figures in Arabic numerals. The title should be short, but expressive. Figures, diagrams and photographs should be embedded to the text. The title of the figure should be aligned to the centre in italic 10 pt size characters under the figures.

Tables: The same rules are valid for figures and tables. Use tabs instead of spaces or hard returns when setting up columns. In tables do not use vertical lines. Avoid excessive number of digits in the body of the table. Refer to each table in the text. The title of the tables should be aligned to the centre in italic 10 pt size characters above the tables.

More information on publication may be obtained from the Editorial Office:

Dr. habil Angéla Anda

Hungarian University of Agriculture and Life Sciences, Georgikon Campus

Tel: +36 83/545-149

E-mail: anda.angela@uni-mate.hu; anda@keszthelynet.hu